

Prenatal Skeletal Dysplasia Panel

Disorder also known as: Osteochondrodysplasias

Panel Gene List: AGPS, ALPL, ARSE, BMP1, CEP120, COL11A1, COL11A2, COL1A1, COL1A2, COL2A1, COMP, CRTAP, DLL3, DYNC2H1, EBP, EVC, EVC2, FGFR1, FGFR2, FGFR3, FKBP10, FLNA, FLNB, GNPAT, HSPG2, IFITM5, IFT172, INPPL1, KIAA0586, LBR, LEPRE1, LIFR, NEK1, PEX7, PLOD2, POR, PPIB, RUNX2, SERPINH1, SLC26A2, SLC35D1, SOX9, TMEM38B, TRIP11, TRPV4, TTC21B, WDR34, WDR35

Clinical Features:

Skeletal dysplasias are a highly variable group of disorders affecting the bone and cartilage of the skeletal system, which are estimated to occur in 2.4 to 4.5 per 10,000 births and 20 per 10,000 stillbirths.¹⁻³ They are characterized by generalized structural abnormalities of bone and cartilage growth and modeling caused by a disturbance in bone growth beginning in the early stages of fetal development and evolving throughout life.³⁻⁶ There are over 450 currently recognized skeletal dysplasias, which are divided into 40 categories based on molecular, biochemical and radiographic criteria.¹⁻³ Although each disorder presents with its own clinical findings, as a group, these conditions are characterized by anomalies of bone shape, size and density, which manifest as abnormalities of the limbs, chest, or skull.⁸ These conditions have variable etiologies including, chromosomal abnormalities or single-gene pathogenic variants as well as environmental factors such as teratogen exposure and autoimmune response.³

While there are a large number of different skeletal dysplasias, certain disorders are more common than others. A brief overview of some of the more common fetal skeletal dysplasias is given below:

Chondrodysplasia Punctata is a group of disorders characterized by chondrodysplasia punctata (stippled epiphyses). The most common form, rhizomelic chondrodysplasia punctata type 1 (RCDP1), is caused by pathogenic variants in the PEX7 gene and is a peroxisome biogenesis disorder characterized by proximal shortening of the humerus and femur, punctate calcifications in cartilage with epiphyseal and metaphyseal abnormalities, congenital cataracts, low birth weight, length, and head circumference, severe postnatal growth deficiency, profound intellectual disability and seizures.¹⁷ Less common disorders result from pathogenic variants in the GNPAT gene causing RCDP2, AGPS gene pathogenic variants causing RCDP3, ARSE pathogenic variants causing X-linked chondrodysplasia punctata 1 (CDPX1) and EBP pathogenic variants causing X-linked chondrodysplasia punctata 2 (CDPX2). These related disorders have similar punctate cartilaginous changes with variable limb shortening and/or asymmetry, short stature, intellectual disability, cataracts, and skin changes.¹⁷

FGFR3-related skeletal dysplasias refer to four distinct disorders caused by pathogenic variants in the FGFR3 gene. The most common of these is achondroplasia (ACH), which is nonlethal and the most common condition associated with disproportionate short stature or dwarfism.^{5,7} Prenatally, this disorder often presents in the third trimester and is associated with rhizomelic micromelia, macrocephaly with frontal bossing and midface hypoplasia. Mild limb bowing, brachydactyly, increased space between the third and fourth digits, and a depressed nasal bridge are also common.^{3,5-7} ACH is estimated to occur in 1 in 10,000 to 1 in 40,000 births with more than 250,000 affected individuals worldwide.^{5,7} Hypochondroplasia (HCH) has a similar, but milder, phenotype to that of ACH and presents with micromelia, short stature and lumbar lordosis.^{5,7} The prevalence of HCH is estimated to be 1 in 50,000 births, and together ACH and HCH are estimated to account for 20% of all cases of skeletal dysplasia in live births.⁵ Thanatophoric dysplasia (TD) is the most common lethal skeletal dysplasia and has an incidence estimated to be between 1 in 17,000 and 1 in 50,000 births.^{3,5} This disorder is characterized by disproportionate dwarfism with very short extremities, normal trunk length, very narrow thorax, macrocephaly, depressed nasal bridge, prominent forehead with protruding eyes, brachydactyly, platyspondyly, and normal bone mineralization without fractures.³⁻⁶ Severe achondroplasia with developmental delay and acanthosis nigricans (SADDAN) is a very severe form of achondroplasia caused by a rare pathogenic variant in the FGFR3 gene.^{5,7}

Osteogenesis Imperfecta (OI) is characterized by bone fragility and consequent susceptibility to bone fractures. The severity of OI can range from severe perinatal lethal to asymptomatic with mild predisposition to fractures and a normal lifespan.^{1,4,8} Other common characteristics include dentinogenesis imperfecta, blue sclerae, short stature and hearing loss in adulthood.⁸ The most lethal form of OI is type II, which is characterized by compressible thin calvaria, severe micromelia and bowing of long bones with multiple fractures and a narrow thorax.³ Together, all types of OI have a combined prevalence of between 1 in 15,000 and 1 in 30,000 births with about 90% of cases caused by pathogenic variants in either COL1A1 or COL1A2.^{4,8}

Achondrogenesis is a severe skeletal dysplasia classified into three types: type IA, type IB, and type II and characterized by a lack of ossification of the vertebral bodies as well as extreme micromelia, a barrel-shaped short trunk, and short ribs.^{3,6} The most common Type II accounts for approximately 80% of cases of achondrogenesis and is due to de novo dominant pathogenic variants in the COL2A1 gene.^{3,6} Type 1A is due to pathogenic variants in the DTDST (SLC26A2) gene, and type IB is due to pathogenic variants in the TRIP11 gene.^{3,10} All three types are usually lethal in the perinatal period.⁶

Short-rib thoracic dysplasias (SRTDs) are characterized by a constricted thoracic cage, short ribs, shortened tubular bones, and a 'trident' appearance of the acetabular roof (lateral

surface of the hip bone). They are autosomal recessive and lethal. All are a part of a group of skeletal ciliopathies caused by problems with cilia and all involve bone abnormalities. Short-rib thoracic dysplasia-6 with or without polydactyly (SRTD6) is caused by pathogenic variants in the NEK1 gene and short-rib thoracic dysplasia-3 with or without polydactyly (SRTD3) is caused by pathogenic variants in the DYNC2H1 gene. Ellis-van Creveld syndrome (EVC) is an autosomal recessive condition additionally characterized by disproportionate short stature, congenital heart disease (most commonly ASD), postaxial polydactyly, dysplastic nails and teeth, and retinal degeneration.^{18,19} This disorder, caused by pathogenic variants in EVC and EVC2, may present prenatally with narrow thorax, shortening of the long bones, polydactyly and cardiac defects.²⁰

Campomelic dysplasia (CD) is a rare, often lethal skeletal dysplasia characterized by angular bowing and shortening of the long bones, severe respiratory distress, and XY sex reversal. It is caused by chromosome abnormalities or pathogenic variants affecting expression of the SOX9 gene located on chromosome 17q24.3-q25.1. Approximately 75% of patients with CD with a 46, XY karyotype exhibit partial or complete sex reversal, ranging from ambiguous genitalia to normal female external genitalia.¹¹ In addition to bowing of the long bones, skeletal features of CD include club feet, a bell-shaped and underdeveloped thorax, eleven pairs of ribs, and hypoplastic scapulae. Other variable features include micrognathia and Pierre-Robin malformation. Many infants die shortly after birth from respiratory compromise; however, those who survive the neonatal period can develop hearing loss, developmental delay, short stature and progressive kyphoscoliosis.¹²

Prenatal Ultrasound Findings:

Skeletal dysplasias are commonly identified in the prenatal period by the presence of shortened long bones or other abnormal skeletal findings such as narrow thorax, polydactyly, frontal bossing, or poor mineralization of the calvarium in an ultrasound.^{3,5} Other imaging methods, such as 3D ultrasound, MRI and CT scan, can be used to evaluate and diagnose a skeletal dysplasia in utero.³ Due to genetic heterogeneity and overlapping phenotypes, the specific fetal skeletal dysplasia cannot be determined accurately with imaging alone.³⁻⁵ When available, molecular, genetic and/or biochemical testing can aid in determining the precise diagnosis after the differential has been established by imaging.

Inheritance Pattern/Genetics:

Many severe skeletal dysplasias are due to single-gene disorders inherited in an autosomal dominant manner and are often sporadic pathogenic variants. Autosomal recessive and X-linked inheritance patterns are also observed.^{1,3,9}

Test Methods:

Using genomic DNA extracted from the submitted specimen, the complete coding regions and splice site junctions of the genes tested are enriched using a proprietary targeted capture system developed by GeneDx for next-generation sequencing with CNV calling (NGS-CNV). The enriched targets are simultaneously sequenced with paired-end reads on an Illumina platform. Bi-directional sequence reads are assembled and aligned to reference sequences based on NCBI RefSeq transcripts and human genome build GRCh37/UCSC hg19. After gene specific filtering, data are analyzed to identify sequence variants and most deletions and duplications involving coding exons; however, technical limitations and inherent sequence properties effectively reduce this resolution for some genes. Alternative sequencing or copy number detection methods are used to analyze or confirm regions with inadequate sequence or copy number data by NGS. Reportable variants include pathogenic variants, likely pathogenic variants and variants of uncertain significance. Likely benign and benign variants, if present, are not routinely reported but are available upon request.

The technical sensitivity of sequencing is estimated to be >99% at detecting single nucleotide events. It will not reliably detect deletions greater than 20 base pairs, insertions or rearrangements greater than 10 base pairs, or low-level mosaicism. The copy number assessment methods used with this test cannot reliably detect copy number variants of less than 500 base pairs or mosaicism and cannot identify balanced chromosome aberrations. Assessment of exon-level copy number events is dependent on the inherent sequence properties of the targeted regions, including shared homology and exon size.

Additionally, genotype analysis of maternal and fetal DNA for several polymorphic markers to test for maternal cell contamination will be performed. Therefore, in all prenatal cases a maternal sample should accompany the fetal sample

Test Sensitivity:

Skeletal dysplasias are a genetically heterogeneous group of disorders with a wide pathogenic variant spectrum. The sensitivity of sequence analysis of this panel in prenatal cases ascertained based on fetal ultrasound abnormalities is currently unknown, and the clinical sensitivity of analysis of the 23 genes included in the Prenatal Skeletal Dysplasias Panel depends on the clinical phenotype of the patient. Specific information about the sensitivity of each gene in selected populations is included in the attached clinical sensitivity table.

Gene	Inheritance	Disease Associations	Diagnostic Yield for Disorder
<i>AGPS</i>	Autosomal recessive	Rhizomelic chondrodysplasia punctata type 3	Unknown
<i>ALPL</i>			
<i>ARSE</i>	X-Linked recessive	Chondrodysplasia punctata	60-75% for sequence variants, multi-exonic and whole-gene deletions in affected males ¹
<i>COL1A1</i>	Autosomal	Osteogenesis imperfecta, types I, II, III & IV	>95% ²

Gene	Inheritance	Disease Associations	Diagnostic Yield for Disorder
	dominant		
<i>COL1A2</i>	Autosomal dominant, Autosomal recessive	Osteogenesis imperfecta, types II, III & IV	>95% ²
<i>COL2A1</i>	Autosomal dominant	Achondrogenesis, type II (ACH2) Hypochondrogenesis Spondyloepiphyseal dysplasia (SED) with metatarsal shortening (also called Czech dysplasia) Spondyloepiphyseal dysplasia (SED) congenita Spondyloepiphyseal dysplasia (SED) (Namaqualand type) Spondyloepimetaphyseal (SMED) (Strudwick type) Otospondylomegaepiphyseal dysplasia Spondyloperipheral dysplasia Platyspondylic skeletal dysplasia (Torrance type) Kniest dysplasia	>75% of COL2A1-related disorders ³
<i>CRTAP</i>	Autosomal recessive	Osteogenesis Imperfecta, type IIB and VII	
<i>DLL3</i>	Autosomal recessive	Spondylocostal dysostosis type 1	
<i>DYNC2H1</i>	Autosomal recessive	Asphyxiating thoracic dystrophy 3	Unknown
<i>EBP</i>	X-Linked dominant	Chondrodysplasia punctata	~70% ⁴
<i>EVC</i>	Autosomal recessive	Ellis-Van Creveld Syndrome	Unknown
<i>EVC2</i>	Autosomal recessive	Ellis-van Creveld syndrome	~90% in affected females ⁵
<i>FGFR2</i>	Autosomal dominant	Bent bone dysplasia	~74% ⁶
<i>FGFR3</i>	Autosomal dominant	Achondroplasia Thanatophoric dysplasia, type I / II Hypocondroplasia	~26% ⁶

Gene	Inheritance	Disease Associations	Diagnostic Yield for Disorder
<i>FLNB</i>	Autosomal dominant	Atelosteogenesis, type I / III (AOI / AOIII) Boomerang dysplasia (BD) Larsen syndrome Spondylocarpotarsal synostosis syndrome (SCT) *Autosomal recessive	Unknown
<i>GNPAT</i>	Autosomal recessive	Rhizomelic chondrodysplasia punctata type 2	>99% ^{7,8,9}
<i>IFITM5</i>	Autosomal dominant	Osteogenesis imperfecta type 5	
<i>LEPRE1</i>	Autosomal recessive	Osteogenesis imperfecta type VIII	
<i>NEK1</i>	Autosomal recessive	Short rib-polydactyly syndrome, Majewski type Asphyxiating thoracic dystrophy 1	~97% in patients with a radiographic diagnosis ¹⁰
<i>PEX7</i>	Autosomal recessive	Rhizomelic chondrodysplasia punctata type 1	
<i>PPIB</i>	Autosomal recessive	Osteogenesis imperfecta, type IX	Unknown
<i>SLC26A2</i> (<i>DTDST</i>)	Autosomal recessive	Achondrogenesis type 1B Atelosteogenesis type II Diastrophic dysplasia	Unknown
<i>SOX9</i>	Autosomal dominant	Campomelic dysplasia	Unknown
<i>TRIP11</i>	Autosomal recessive	Achondrogenesis, type IA	Unknown

Table References

- Braverman NE, Bober M, Brunetti-Pierri N, et al. Chondrodysplasia Punctata 1, X-Linked. 2008 Apr 22 [Updated 2014 Nov 20]. In: Pagon RA, Adam MP, Ardinger HH, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2016. Available from: <http://www.ncbi.nlm.nih.gov/books/NBK1544/>.
- Steiner RD, Adsit J, Basel D. COL1A1/2-Related Osteogenesis Imperfecta. 2005 Jan 28 [Updated 2013 Feb 14]. In: Pagon RA, Adam MP, Ardinger HH, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2016. Available from: <http://www.ncbi.nlm.nih.gov/books/NBK1295/>.
- Nishimura G et al. Human Mutation. 2005 Jul 26(1):36-43.15895462.
- Nishimura G et al. Human Mutation. 2005 Jul 26(1):36-43.15895462.
- Dempsey MA, Tan C, Herman GE. Chondrodysplasia Punctata 2, X-Linked. 2011 May 31. In: Pagon RA, Adam MP, Ardinger HH, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2016. Available from: <http://www.ncbi.nlm.nih.gov/books/NBK55062/>.
- D'Asdia MC et al. European Journal Of Medical Genetics. 2013 56(2):80-7.23220543.
- Pauli RM. Achondroplasia. 1998 Oct 12 [Updated 2012 Feb 16]. In: Pagon RA, Adam MP, Ardinger HH, et al.,

- editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2016. Available from: <http://www.ncbi.nlm.nih.gov/books/NBK1152/>.
8. Karczeski B, Cutting GR. Thanatophoric Dysplasia. 2004 May 21 [Updated 2013 Sep 12]. In: Pagon RA, Adam MP, Ardinger HH, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2016. Available from: <http://www.ncbi.nlm.nih.gov/books/NBK1366/>
 9. Bober MB, Bellus GA, Nikkel SM, et al. Hypochondroplasia. 1999 Jul 15 [Updated 2013 Sep 26]. In: Pagon RA, Adam MP, Ardinger HH, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2016. Available from: <http://www.ncbi.nlm.nih.gov/books/NBK1477/>.
 10. Daniel et al. (2012). "Disease-associated mutations in the actin-binding domain of filamin B cause cytoplasmic focal accumulations correlating with disease severity". *Hum Mutat* 33(4):665-673. PubMed ID: 22190451.
 11. Rossi A, Superti-Furga A. *Hum Mutat*. 2001;17:159–71.; Bonafé L, Mittaz-Crettol L, Ballhausen D, et al. Achondrogenesis Type 1B. 2002 Aug 30 [Updated 2013 Nov 14]. In: Pagon RA, Adam MP, Ardinger HH, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2016. Available from: <http://www.ncbi.nlm.nih.gov/books/NBK1516/>.
 12. Bonafé L, Mittaz-Crettol L, Ballhausen D, et al. Atelosteogenesis Type 2. 2002 Aug 30 [Updated 2014 Jan 23]. In: Pagon RA, Adam MP, Ardinger HH, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2016. Available from: <http://www.ncbi.nlm.nih.gov/books/NBK1317/>.
 13. Bonafé L, Mittaz-Crettol L, Ballhausen D, et al. Achondrogenesis Type 1B. 2002 Aug 30 [Updated 2013 Nov 14]. In: Pagon RA, Adam MP, Ardinger HH, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2016. Available from: <http://www.ncbi.nlm.nih.gov/books/NBK1516/>.
 14. Pop R et al. A homozygous nonsense mutation in SOX9 in the dominant disorder campomelic dysplasia: a case of mitotic gene conversion. *Human Genetics*. 2005 117(1):43-53.; Unger S, Scherer G, Superti-Furga A. Campomelic Dysplasia. 2008 Jul 31 [Updated 2013 May 9]. In: Pagon RA, Adam MP, Ardinger HH, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2016. Available from: <http://www.ncbi.nlm.nih.gov/books/NBK1760/>

References:

1. Barkova, E et al. (2014). *Clinical Genetics*.doi:10.1111/cge.12434 [doi]
2. Nelson, DB et al. (2014). *Journal of Ultrasound in Medicine* 33(6), 1085-1090.
3. Noel, AE & Brown RN (2014). *International Journal of Women's Health*, 6, 489-500.
4. Dighe, M, et al. (2008). *Radiographics* 28(4), 1061-1077.
5. Hatzaki, A et al. (2011). *American Journal of Medical Genetics*, 155A(10), 2426-2435.
6. Witters, I, Moerman, P & Fryns, JP (2008). *Genetic Counseling (Geneva, Switzerland)*, 19(3), 267-275.
7. Stratbucker, WB (2009). *Pediatrics in Review*, 30(3), 114-115.
8. Valadares, ER, et. al (2014). *Jornal De Pediatria*, 90(6), 536-541.
9. Krakow, D, et. al., (2009). *Genetics in Medicine* 11(2), 127-133.
10. <http://www.omim.org/entry/200600>
11. Mansour et al., (1995) *J Med Genet* 32:415-420.
12. Mansour et al., (2002) *J Med Genet* 39:597-602.
13. Pop et al. (2005) *Hum Genet* 117:43-53.
14. Moog et al., (2001) *Am J Med Genet* 104:239-245.
15. Pop et al., (2004) *J Med Genet* 41:e47.
16. Meyer et al., (1997) *Hum Mol Genet* 6(1):91-98
17. Braverman, N, Moser, A and Steinberg, S. (Updated September 13, 2012). Rhizomelic Chondrodysplasia Punctata Type 1. In: GeneReviews at GeneTests Medical Genetics Information Resource (database online). Copyright, University of Washington, Seattle. 1997-2015. Available at <http://www.genetests.org>. Accessed [Jan 2016].
18. Gunay-Aygun, M, Gahl,W, and Heller,T (Updated April 24, 2014). Congenital Hepatic Fibrosis Overview. In: GeneReviews at GeneTests Medical Genetics Information Resource (database online). Copyright, University of Washington, Seattle. 1997-2015. Available at <http://www.genetests.org>. Accessed [Jan 2016].
19. Peraita-Ezcurra M, et al. (2012) *Gene*. May 10;499(1):223-5.
20. Baujat G, et al. (2007) *Orphanet J Rare Dis*. Jun 4;2:27.
21. Bober, M. B., Taylor, M., Heinle, R., & Mackenzie, W. (2012). Achondroplasia-hypochondroplasia complex and abnormal pulmonary anatomy. *American Journal of Medical Genetics*.Part A, 158A(9), 2336-2341. doi:10.1002/ajmg.a.35530 [doi]. Drera, B., Ferrari, D., Cavalli, P., & Poggiani, C. (2014). A case of neonatal jeune syndrome expanding the phenotype. *Clinical Case Reports*, 2(4), 156-158. doi:10.1002/ccr3.85 [doi]
22. Honorio, J. C., Bruns, R. F., Grundtner, L. F., Raskin, S., Ferrari, L. P., Araujo Junior, E., & Nardoza, L. M. (2013). Diastrophic dysplasia: Prenatal diagnosis and review of the literature. *Sao Paulo Medical Journal = Revista Paulista De Medicina*, 131(2), 127-132. doi:S1516-31802013000200127 [pii]
23. Khalil, A., Pajkrt, E., & Chitty, L. S. (2011). Early prenatal diagnosis of skeletal anomalies. *Prenatal Diagnosis*, 31(1), 115-124. doi:10.1002/pd.2676 [doi]
24. Nampoothiri, S., Yesodharan, D., Sainulabdin, G., Narayanan, D., Padmanabhan, L., Girisha, K. M., Superti-Furga, A. (2014). Eight years experience from a skeletal dysplasia referral center in a tertiary hospital in southern india: A model for the diagnosis and

treatment of rare diseases in a developing country. *American Journal of Medical Genetics. Part A*, 164A(9), 2317-2323. doi:10.1002/ajmg.a.36668 [doi]

25. Miyazaki, O., Nishimura, G., Sago, H., Horiuchi, T., Hayashi, S., & Kosaki, R. (2012). Prenatal diagnosis of fetal skeletal dysplasia with 3D CT. *Pediatric Radiology*, 42(7), 842-852. doi:10.1007/s00247-012-2381-7 [doi]
26. Rice, K. J., Ballas, J., Lai, E., Hartney, C., Jones, M. C., & Pretorius, D. H. (2011). Diagnosis of fetal limb abnormalities before 15 weeks: Cause for concern. *Journal of Ultrasound in Medicine : Official Journal of the American Institute of Ultrasound in Medicine*, 30(7), 1009-1019. doi:30/7/1009 [pii]
27. Yeh, P., Saeed, F., Paramasivam, G., Wyatt-Ashmead, J., & Kumar, S. (2011). Accuracy of prenatal diagnosis and prediction of lethality for fetal skeletal dysplasias. *Prenatal Diagnosis*, 31(05), 515-518. doi:10.1002/pd.2729 [doi]