

Prenatal Lissencephaly Panel Sequence Analysis and Exon-Level Deletion/Duplication Testing of 26 Genes

Panel Gene List: ACTB*, ACTG1, ARX*, ATP6V0A2, B3GALNT2, B4GAT1**, CIT, DCX, FKRP**, FKTN, GMPPB, ISPD, KATNB1, LAMB1, LARGE1, NDE1, PAFAH1B1, POMGNT1*, POMGNT2, POMT1, POMT2, RELN, TMEM5, TUBA1A*, VLDLR, WDR62

*Only large deletion/duplications may be detected for the ACTB, ARX, POMGNT1 and TUBA1A genes

**No deletion/duplication analysis for the B4GAT1 and FKRP genes

Clinical Features:

Lissencephaly is characterized by a thickened cortex and the absence of folds or gyri (agyria) or the presence of abnormally wide gyri (pachygyria). Lissencephalies can present in different forms and with varying severity, ranging from complete agyria to mixed agyria and pachygyria, or simplified gyri with subcortical band heterotopia (double cortex). Pathogenic variants in distinct genes lead to overlapping forms of lissencephaly, including classic smooth lissencephaly, cobblestone lissencephaly, lissencephaly with agenesis of the corpus callosum, and lissencephaly with cerebellar hypoplasia. Subcortical band heterotopia is a mild form of lissencephaly characterized by normal gyri but the presence of an abnormal and often symmetric band of gray matter under the cortex. Lissencephalies uniformly cause developmental delay, epilepsy, and intellectual disability.¹ The spectrum of disorders associated with lissencephaly includes a group of congenital muscular dystrophies called the alpha-dystroglycanopathies including Walker-Warburg syndrome which is characterized by cobblestone lissencephaly, cerebellar abnormalities, eye defects, muscle weakness.^{2,3}

Lissencephaly may not be seen on ultrasound before 24 weeks of gestation.⁴ Other imaging methods, such as 3D ultrasound or fetal MRI, may aid in the diagnosis of lissencephaly in utero.^{4,5,6} Due to genetic heterogeneity and overlapping phenotypes, the specific diagnosis cannot be determined accurately with imaging alone. When available, genetic testing can aid in determining the precise diagnosis after the differential has been established by imaging.

Inheritance Pattern/Genetics:

The various forms of lissencephaly demonstrate clinical and genetic heterogeneity.^{1,2} Classic lissencephaly occurs as an autosomal dominant, autosomal recessive, or X-linked trait. PAFAH1B1 (LIS1) is among the most prominent genes associated with lissencephaly. Loss of PAFAH1B1 alone causes epilepsy and developmental delay, but with absence of severe dysmorphism. Deletion of this gene along with adjacent ones in the terminal end of the short arm of chromosome 17 causes Miller-Dieker syndrome, characterized by epilepsy, dysmorphic features, severe developmental delay and, occasionally, heart defects and omphalocele. Somatic mutations in PAFAH1B1 and DCX have been described.^{7,8} Cobblestone lissencephaly

and lissencephaly with cerebellar hypoplasia segregate as autosomal recessive disorders and lissencephaly with agenesis of the corpus callosum is an X-linked disorder. Lissencephaly can sometimes occur together with heterotopias or polymicrogyria, depending on the affected gene and the type of pathogenic variant.

Test Methods:

Using genomic DNA from the submitted specimen, the complete coding regions and splice site junctions of the genes on this panel are enriched using a proprietary targeted capture system developed by GeneDx for next-generation sequencing with CNV calling (NGS-CNV). The enriched targets are simultaneously sequenced with paired-end reads on an Illumina platform. Bi-directional sequence reads are assembled and aligned to reference sequences based on NCBI RefSeq transcripts and human genome build GRCh37/UCSC hg19. After gene specific filtering, data are analyzed to identify sequence variants and most deletions and duplications involving coding exons. Alternative sequencing or copy number detection methods are used to analyze regions with inadequate sequence or copy number data. Reportable variants include pathogenic variants, likely pathogenic variants and variants of uncertain significance. Likely benign and benign variants, if present, are not routinely reported but are available upon request.

Additionally, genotype analysis of maternal and fetal DNA for several polymorphic markers to test for maternal cell contamination will be performed. Therefore, in all prenatal cases a maternal sample should accompany the fetal sample.

Test Sensitivity:

The clinical sensitivity of sequencing and deletion/duplication analysis of the genes included in this panel in prenatal cases ascertained based on fetal ultrasound abnormalities is currently unknown, and the clinical sensitivity of analysis of the 24 genes including on the Prenatal Lissencephaly panel depends in part on the patient's clinical phenotype. Specific information about the diagnostic yield for each gene in selected populations is summarized in the following table(s).

The technical sensitivity of sequencing is estimated to be > 99% at detecting single nucleotide events. It will not reliably detect deletions greater than 20 base pairs, insertions or rearrangements greater than 10 base pairs, or low-level mosaicism. The copy number assessment methods used with this test cannot reliably detect copy number variants of less than 500 base pairs or mosaicism and cannot identify balanced chromosome aberrations. Assessment of exon-level copy number events is dependent on the inherent sequence properties of the targeted regions, including shared homology and exon size. Gene specific exclusions for exon-level deletion/duplication testing for this panel are: B4GAT1 and FKRP

genes, no copy number testing, ACTB, ARX, POMGNT1 and TUBA1A genes, only whole gene deletions or duplications may be detected.

Gene	Protein	Inheritance	Disease Associations
<i>ACTB</i> *	Actin, Beta	AD	80% of Baraitser-Winter syndrome ⁹
<i>ACTG1</i>	Actin, Gamma-1	AD	20% of Baraitser-Winter syndrome ⁹
<i>ARX</i> *	Aristaless-related homeobox protein	XL	70-95% of XLAG7,8, 3-10% in XLID ^{12,13}
<i>ATP6V0A2</i>	Lysosomal H(+)-ATPase V0 subunit A2	AR	21-24% of autosomal recessive cutis laxa type II ^{14,15}
<i>B3GALNT2</i>	Beta-1,3-N-Acetylgalactosaminyltransferase 2	AR	Rare in alpha-dystroglycanopathies ¹⁶ Rare in congenital muscular dystrophy ¹⁷
<i>B4GAT1</i> (<i>B3GNT1</i>)**	Beta-1,4-Glucuronyltransferase 1	AR	Rare in alpha-dystroglycanopathies ^{18,19}
<i>CIT</i>	Citron rho-interacting serine/threonine kinase	AR	Rare in MCPH ^{20,21,22}
<i>DCX</i>	Doublecortin	XL	Up to 100% XL lissencephaly 10% of classic lissencephaly 85% females and ~30% males with SBH ^{1,11,23}
<i>FKRP</i> **	Fukutin-related protein	AR	~2% of cobblestone lissencephaly ^{3,24} 9% of alpha-dystroglycanopathies ²⁵ 6% of limb-girdle MD ²⁶ 2.5% of congenital muscular dystrophy ¹⁷
<i>FKTN</i>	Fukutin	AR	~7% of alpha-dystroglycanopathies ²⁷ Rare in congenital muscular dystrophy ¹⁷ 1.5% of limb-girdle MD ²⁸ Does not include the Japanese founder mutation in the 3' UTR ²⁹
<i>GMPPB</i>	GDP-Mannose Pyrophosphorylase B	AR	Rare in alpha-dystroglycanopathies ^{30,31}

			Rare in congenital muscular dystrophy ¹⁷
<i>ISPD</i>	Isoprenoid synthase domain-containing protein	AR	~6% of cobblestone lissencephaly ²⁷ ~30% of Walker-Warburg syndrome and ~11% of alpha-dystroglycanopathies ^{32,33} Rare in LGMD ³⁴
<i>KATNB1</i>	Katanin regulatory subunit B1	AR	<1% of MCD ^{35,36}
<i>LAMB1</i>	Laminin, Beta-1	AR	Rare in cobblestone lissencephaly ³⁷
<i>LARGE1</i>	Like-glycosyltransferase	AR	2-5% of cobblestone lissencephaly ^{3,24} ~1% of alpha-dystroglycanopathies ^{25,27}
<i>NDE1</i>	nudE neurodevelopment protein 1	AR	Rare ^{38,39,40}
<i>PAFAH1B1 (LIS1)</i>	Platelet-Activating Factor Acetylhydrolase 1b, Regulatory Subunit 1	AD	~40-65% of classic lissencephaly ^{1,41}
<i>POMGNT1*</i>	Protein O-Mannose Beta 1-2-N-Acetylglucosaminyltransferase	AR	11-18% of cobblestone lissencephaly ^{3,24} 8-10% of alpha-dystroglycanopathies ^{25,27}
<i>POMGNT2</i>	Protein O-Mannose Beta-1,4-N-Acetylglucosaminyltransferase 2	AR	Rare in alpha-dystroglycanopathies ⁴²
<i>POMT1</i>	Protein O-Mannosyltransferase 1	AR	27-34% of cobblestone lissencephaly ^{3,24} 9-21% of alpha-dystroglycanopathies ^{25,27}
<i>POMT2</i>	Protein O-Mannosyltransferase 2	AR	8-11% of cobblestone lissencephaly ^{3,24} 9-11% of alpha-dystroglycanopathies ^{25,27}
<i>RELN</i>	Reelin	AR	Rare ^{43,44}
<i>TMEM5</i>	Transmembrane protein 5	AR	~6% of cobblestone lissencephaly ²⁴ Rare in alpha-dystroglycanopathies ⁴⁵
<i>TUBA1A*</i>	Tubulin, Alpha-1A	AD	1% of classic lissencephaly 30% of lissencephaly with cerebellar hypoplasia ^{46,47}
<i>VLDLR</i>	Very low density lipoprotein receptor	AR	Rare cerebellar hypoplasia with simplified gyri ^{48,49}

WDR62	WD repeat-containing protein 62	AR	Unknown ⁵⁰
-------	---------------------------------	----	-----------------------

*Only large deletion/duplications may be detected for the ACTB, ARX, POMGNT1 and TUBA1A genes

**No deletion/duplication analysis for the B4GAT1 and FKRP genes

References:

1. Fry et al. (2014) American Journal Of Medical Genetics. Part C, Seminars In Medical Genetics 166C (2):198-210 (PMID: 24862549).
2. Dymont et al. (2013) Current Neurology And Neuroscience Reports 13 (8):364 (PMID: 23793931)
3. Devisme et al. (2012) Brain : A Journal Of Neurology 135 (Pt 2):469-82 (PMID: 22323514)
4. Senapati et al. (2012) J Pediatr Neuroradiol 1 (3):171-184 (PMID: 24078783)
5. Conte et al. (2016) AJNR Am J Neuroradiol 37 (5):946-51 (PMID: 26721771)
6. Williams et al. (2017) Br J Radiol :20160902 (PMID: 28134568)
7. Dobyns WB, Das S. LIS1-Associated Lissencephaly/Subcortical Band Heterotopia. 2009 Mar 3 [Updated 2014 Aug 14]. In: Pagon RA, Adam MP, Ardinger HH, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2015. Available from: <http://www.ncbi.nlm.nih.gov/books/NBK5189/>
8. Hehr U, Uyanik G, Aigner L, et al. DCX-Related Disorders. 2007 Oct 19 [Updated 2011 Mar 24]. In: Pagon RA, Adam MP, Ardinger HH, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2015. Available from: <http://www.ncbi.nlm.nih.gov/books/NBK1185/>
9. Verloes et al. (2015) European Journal Of Human Genetics : Ejhg 23 (3):292-301 (PMID: 25052316)
10. Kato et al. (2004) Human Mutation 23 (2):147-59 (PMID: 14722918)
11. Dobyns (2010) Epilepsia 51 Suppl 1 :5-9 (PMID: 20331703)
12. Poirier et al. (2006) Neurogenetics 7 (1):39-46 (PMID: 16235064)
13. Nawara et al. (2006) American Journal Of Medical Genetics. Part A 140 (7):727-32 (PMID: 16523516)
14. Gardeitchik et al. (2014) European Journal Of Human Genetics : Ejhg 22 (7):888-95 (PMID: 23963297)
15. Fischer et al. (2012) Human Genetics 131 (11):1761-73 (PMID: 22773132)
16. Stevens et al. (2013) American Journal Of Human Genetics 92 (3):354-65 (PMID: 23453667)
17. Sframeli et al. (2017) Neuromuscul. Disord. 27 (9):793-803 (PMID: 28688748)
18. Buysse et al. (2013) Human Molecular Genetics 22 (9):1746-54 (PMID: 23359570)
19. Shaheen et al. (2013) Neurogenetics 14 (3-4):243-5 (PMID: 23877401)
20. Li et al. (2016) Am. J. Hum. Genet. 99 (2):501-10 (PMID: 27453578)
21. Basit et al. (2016) Hum. Genet. 135 (10):1199-207 (PMID: 27519304)
22. Harding et al. (2016) Am. J. Hum. Genet. 99 (2):511-20 (PMID: 27453579)
23. Bahi-Buisson et al. (2013) Brain : A Journal Of Neurology 136 (Pt 1):223-44 (PMID: 23365099)
24. Vuillaumier-Barrot et al. (2012) American Journal Of Human Genetics 91 (6):1135-43 (PMID: 23217329)
25. Mercuri et al. (2009) Neurology 72 (21):1802-9 (PMID: 19299310)
26. Pegoraro E, Hoffman EP. Limb-Girdle Muscular Dystrophy Overview. 2000 Jun 8 [Updated 2012 Aug 30]. In: Pagon RA, Adam MP, Ardinger HH, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2015. Available from: <http://www.ncbi.nlm.nih.gov/books/NBK1408/>
27. Godfrey et al. (2007) Brain : A Journal Of Neurology 130 (Pt 10):2725-35 (PMID: 17878207)
28. Oestergaard et al. (2016) Neurol Genet 2 (6):e112 (PMID: 27766311)
29. Saito K. Fukuyama Congenital Muscular Dystrophy. 2006 Jan 26 [Updated 2012 May 10]. In: Pagon RA, Adam MP, Ardinger HH, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2015. Available from: <http://www.ncbi.nlm.nih.gov/books/NBK1206/>
30. Jensen et al. (2015) Hum. Mutat. : (PMID: 26310427)
31. Carss et al. (2013) American Journal Of Human Genetics 93 (1):29-41 (PMID: 23768512)
32. Czeschik et al. (2013) European Journal Of Medical Genetics 56 (12):689-94 (PMID: 24120487)
33. Willer et al. (2012) Nature Genetics 44 (5):575-80 (PMID: 22522420)
34. Cirak et al. (2013) Brain : A Journal Of Neurology 136 (Pt 1):269-81 (PMID: 23288328)
35. Mishra-Gorur et al. (2014) Neuron 84 (6):1226-39 (PMID: 25521378)
36. Hu et al. (2014) Neuron 84 (6):1240-57 (PMID: 25521379)
37. Radmanesh et al. (2013) American Journal Of Human Genetics 92 (3):468-74 (PMID: 23472759)
38. Bakircioglu et al. (2011) American Journal Of Human Genetics 88 (5):523-35 (PMID: 21529752)
39. Alkuraya et al. (2011) American Journal Of Human Genetics 88 (5):536-47 (PMID: 21529751)
40. Guven et al. (2012) Neurogenetics 13 (3):189-94 (PMID: 22526350)
41. Saillour et al. (2009) Archives Of Neurology 66 (8):1007-15 (PMID: 19667223)
42. Manzini et al. (2012) American Journal Of Human Genetics 91 (3):541-7 (PMID: 22958903)
43. Hong et al. (2000) Nature Genetics 26 (1):93-6 (PMID: 10973257)
44. Zaki et al. (2007) American Journal Of Medical Genetics. Part A 143A (9):939-44 (PMID: 17431900)
45. Jae et al. (2013) Science (New York, N.Y.) 340 (6131):479-83 (PMID: 23519211)

46. Kumar et al. (2010) *Human Molecular Genetics* 19 (14):2817-27 (PMID: 20466733)
47. Cushion et al. (2013) *Brain : A Journal Of Neurology* 136 (Pt 2):536-48 (PMID: 23361065)
48. Ozcelik et al. (2008) *Proceedings Of The National Academy Of Sciences Of The United States Of America* 105 (11):4232-6 (PMID: 18326629)
49. Boycott et al. (2009) *Journal Of Child Neurology* 24 (10):1310-5 (PMID: 19332571)
50. Nicholas et al. (2010) *Nature Genetics* 42 (11):1010-4 (PMID: 20890279)